Chemical Constituents of *Moringa oleifera* Leaves and Seeds from Abakaliki, Nigeria

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ABSTRACT

Moringa oleifera is a medicinal plant widely used in folkloric medicine of Africa and Asia for the treatment of ailments such as ulcer, wound, inflammation, heart problem, cancer, stroke, obesity, anaemia and liver damage. The chemical constituents of the methanolic extract of Moringa oleifera leaves and seeds were investigated using Gas chromatography-mass spectrometry. Sixteen chemical constituents were identified in the leaf methanolic extract; they are 9-octadecenoic acid (20.89%), L-(+)-ascorbic acid- 2,6dihexadecanoate(19.66%), 14-methyl-8-hexadecenal (8.11%), 4hydroxyl-4-methyl-2-pentanone (7.01%), 3-ethyl-2, 4-dimethylpentane (6.14%), phytol (4.24%), octadecamethyl-cyclononasiloxane (1.23%), 1, 2-benzene dicarboxylic acid (2.46%), 3, 4-epoxyethanone comprising (1.78%), N-(-1-methylethyllidene)-benzene ethanamine (1.54%), 4, 8, 12, 16-tetramethylheptadecan-4-olide (2.77%), 3-5-bis (1, 1-dimethylethyl)-phenol (2.55%), 1-hexadecanol (1.23%), 3, 7, 11, 15-tetramethyl-2 hexadecene-1-ol (1.17%), hexadecanoic acid (2.03%) and 1, 2, 3-propanetriyl ester-9 octadecenoic acid(1.23%). Five chemical constituents were identified in methanolic seed extract and they are oleic acid (84%), L-(+) ascorbic acid- 2, 6-dihexadecanoate (9.80%), 9-octadecenoic acid ester-hexadecanoic acid (1.31%) methyl (1.88%). octadecenamide (0.78%). Results obtained showed that the methanolic leaf extract of Moringa oleifera has more chemical constituents than the seed with 9-octadecenoic acid (20.8%) as the highest in the leaf and oleic acid (84%) in the seed. These relatively diverse chemical constituents may be responsible for the medicinal properties of *Moringa oleifera* leaves and seeds.

Keywords: GC-MS analysis, Chemical constituents, Methanol extract, *Moringa oleifera*.

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INTRODUCTION

Medicinal plants have been used by all civilizations as a source of medicines since ancient times⁶. In recent times, there has been growing interest in exploiting the biological activities of different ayurvedic medicinal herbs, due to their natural origin, cost effectiveness and lesser side effects⁶. Interest in medicinal plants as a re-emerging health aid in the maintenance of personal health and well-being has been fuelled by rising costs of prescription drugs and the bio-prospecting of new plant-derived drugs¹⁹. Generally, plants which produce constituents mostly as secondary metabolites medicinal having values are called medicinal plants. These substances differ from plant to plant, thus the plant kingdom provides a large store of various chemical substances with potential therapeutic properties which have been utilized in treatment and cure of human and other animal diseases including relieving of pains, convulsion and cardiovascular diseases¹².

Drugs of natural origin are considered to be less toxic and free from adverse effects than synthetic ones. Even though active compounds of many herbal drugs were unknown, they have been widely prescribed by the practitioners of the traditional medicines due to their minimal adverse effects and low cost¹⁷. World Health Organisation (WHO) has estimated that 4 billion people (80% of the world population) use herbal medicines for some aspect of primary health care²⁰.

Moringa oleifera is commonly known as drumstick-tree or horse radishtree. It is used as vegetable and also in Indian folk medicine for the treatment of various illnesses¹⁴. Moringa oleifera is a small graceful tree with sparse foliage often planted in compounds or used in fencing in Nigeria. It resembles a leguminous species at a distance especially when flowering.

Moringa oleifera is known with the following local names: "Zogallagandi" (Hausa), "Ewe-igbale" (Yoruba) and "Okwe Oyibo" (Igbo)⁹. Moringa oleifera is a common vegetable in Nigeria especially in the Eastern Nigeria. The pictures of Moringa oleifera leaves and seeds are shown in figure 1 and 2 below.

However, apart from its traditional, medicinal and nutritional uses, there are several reports on the biological and physiological activities of *Moringa oleifera*. These include hypotensive properties⁸, hypoglycemic and hypocholesterolemic effects^{4,5,7}, anti-inflammatory and activities¹⁵, anti-helmic. hepatotoxic analgesic, dyspepsia and in the management of heart diseases, and ulcers8. According to Aja et al., the seed of Moringa oleifera revealed that administration of aqueous, ethanolic and methanolic seed extracts of Moringa oleifera significantly reduced (p<0.05) the levels of some liver enzymes in albino rats. They also observed that the liver pathology showed that no significant lesions were observed and this may point to the fact that the seed of *Moringa oleifera* is relatively safe for use medicinally.

Despite the popular use of *Moringa* oleifera leaves and seeds for treating various disorders, there is limited data available regarding Gas chromatography–mass spectrometry (GC/MS) analysis of chemical constituents of locally grown *Moringa* oleifera leaves and seeds in Abakaliki, Nigeria. This study therefore intends to evaluate the Gas chromatography–mass spectrometry (GC/MS) analysis of the chemical constituents of the methanolic extract of *Moringa* oleifera leaves and seeds grown in Abakaliki, Nigeria.

MATERIALS AND METHODS

Plant Collection

The fresh leaves and seeds of *Moringa oleifera* were collected from Abakaliki Area of Ebonyi State, Nigeria and were identified by taxonomist in the Department of Applied Biology, Ebonyi State University, Abakaliki, Nigeria. A part was also deposited in the hebarium for reference purposes.

Preparation of Samples

The leaves were destalked, washed and shade dried at ambient temperature with constant turning averts fungal growth. The seeds were dehusked and dried through the same process. The leaves and seeds were later milled to obtain the vegetable leaf meals (VLMs) and seed meals (SMs) using an electric blender and both were stored in 4°C temperature in refrigerator in well labeled airtight containers for analysis.

Preparation of Extract

40gms of dried powdered leaves and seeds of *Moringa oleifera* were extracted successively with 300 ml of methanol in an orbital shaker for 24 hrs at room temperature. The extracts were filtered using Whatman No.1 filter paper to remove extractable substances, at every 3 hrs interval. The combined extracts were then evaporated with rotary evaporator and the dried extracts were stored at 4°C in two different sterile containers.

GC-MS Analysis

Procedure

GC-MS analysis of the methanolic extract of *M. oleifera* leaves and seeds were performed using Shimadzu Japan gas chromatography QP2010PLUS with a fused GC column (2010) coated with polymethyl silicon (0.25nm x 50m) and the conditions

were as follows: Temperature programming from 80-200°C held at 80°C for 1 min, rate 5°C/min and at 200°c for 20 min. Field ionization detector (FID) temperature 300°C, injection temperature 220°c, carrier gas nitrogen at a flow rate of 1 ml/min, split ratio 1:75. Gas chromatography mass spectrum was conducted using GCMS -QP 2010 Plus Shimadzu Japan with injector temperature of 220°c and carrier gas pressure of 116.9 kpa. The column length is 30 m with a diameter of 0.25 mm and flow rate of 50 ml/min. The elutes were automatically passed into a mass spectrometer with a dictator voltage set at 1.5 ky and sampling rate of 0.2 sec. The mass spectrum was also equipped with a computer fed mass spectra data bank. Hermlez 233 M-Z centrifuge (Germany) was used.

Component Identification

Chemical constituent components of the extracts were identified by matching the peaks with Computer Wiley MS libraries and confirmed by comparing mass spectra of the peaks and those from literature¹¹.

RESULTS

Sixteen peaks were identified from the chromatogram of the methanolic leaf extract of Moringa oleifera (Figure 3). These peaks (1-16) indicate the presence of sixteen compounds (1-16) in the extract (Figure 1). The molecular formula, percentage content and molecular mass of the compounds are shown in Table 1. These compounds comprise mainly hydrocarbons, fatty acids, alcohols, esters and phenols. The composition of the extract comprises; 9-Octadecenoic acid (20.89%), L-(+) - Ascorbic acid- 2, 6dihexadecanoate (19.66%), 14 -methyl -8-Hexadecenal (8.11%), 4- hydroxyl-4-methyl-2-pentanone (7.01%), 3-ethyl-2, 4-dimethylpentane (6.14%) and phytol (4.25%) as the major chemical constituents.

Compound 1 was identified as 4hvdroxyl-4-methyl-2-pentanone molecular formula of C₆H₁₂O₂ (m/z 116) with base peak at m/z 43 which was due to loss of propanone group $((CH_3)_2C=0)$ from the parent molecule. The fragmentation peak at m/z =101 was due to loss of methyl radical while the loss of H₂0 molecule gave weak peak at m/z=83. It constitutes 7.01% of the extract. Compound 2 constitutes 6.14% of the extract with molecular formula C₉H₂₀ (m/z 128) and base peak at m/z 43 which occurred due to the detachment of a propyl fragment C_3H_7 (m/z 43) from the compound. It was identified as 3-ethyl-2, 4-dimethyl-pentane. Compound 3 has molecular formula C₆H₁₀O₂ (m/z 114) and base peak at m/z 43 which was due to the loss of butanone group. The compound was identified as 3, 4-epoxyethanone constituting 1.78% of the extract. Compound 4 is N-(-1-methylethyllidene)benzene ethanamine with molecular formula $C_{11}H_{15}N$ (m/z 161) and base peak at m/z 70 which was due to the loss of benzene methyl group. The constituent was 1.54% of the extract. Compound 5 was identified as 3-5-bis (1, 1-dimethylethyl)-phenol with molecular formula C₁₄H₂₂O (m/z 206) and base peak at m/z 57. The base peak occurred as a result of the detachment of C₃H₉ (m/z 57) fragment from the compound. It constitutes 2.55% of the extract. Compound 6 is 1-hexadecanol with molecular formula C₁₄H₃₄O (m/z 242) and base peak at m/z 55 which was due to loss of propyl group (C₃H₇). It constitutes 1.23% of the extract. Compound 7 was identified as 3, 7, 11, 15-tetramethyl-2 hexadecene-1-ol and with molecular formula $C_{20}H_{40}O$ (m/z 296). It constitutes 1.17% of the extract. Compound 8 was identified as hexadecanoic acid and with molecular formula $C_{17}H_{34}O_2$ (m/z 270) and it constitutes 2.03% of the extract. The base peak occurred as a result of the detachment of C2H5COOH (m/z 74) and hydrogen molecule (H₂) fragments from the compound. Compound 9

has molecular formula $C_{38}H_{68}O_8$ (m/z 652) and comprises 19.66% of the extract. The base peak occurred at C₃H₅O₂ (m/z 73). This peak occurred due to McLafferty rearrangement. Other prominent peaks observed on the compound occurred at m/z 43 ($C_3H_7^+$) and m/z 41 (C₃H₅). These peaks occurred due to proton migration and rearrangement. Compound 9 was identified as L-(+) -Ascorbic acid-2, 6-dihexadecanoate. Compound 10 was identified as phytol with molecular formula C₂₀H₄₀O (m/z 296). It constitutes 4.24% of the extract. The base peak occurred due to loss of methyl butyl group at m/z 71. Compound 11 has a molecular formula of C₅₇H₁₀₄O₆ (m/z 884). It was identified as 9-octadecenoic acid and it constitutes 20.89% of the extract. Compound 12 was identified as 4, 8, 12, 16-tetramethyl heptadecan-4-olide with molecular formula $C_{21}H_{40}O_2$ (m/z 324) and it constitutes 2.77% of the extract. Compound 13 has molecular formula C₁₈H₃₆O₂ (m/z 284) and constitutes 1.23% of the extract. It was identified as 1, 2, 3-propanetriyl ester-9 octadecenoic acid. Compound 14 has molecular formula $C_{17}H_{32}O$ (m/z 252) and it was identified as 14 -methyl -8-hexadecenal. It comprises 8.11% of the extract. Compound 15 has molecular formula $C_{24}H_{38}O_4$ (m/z 390) and was identified as 1, 2-benzenedicarboxylic acid. The base peak occurred at C₄H₉ (m/z 57). It constitutes 2.46% of the extract. Compound 16 has molecular formula C₁₈H₅₄O₉Si₉ (m/z 666) and was identified as octadea methylcyclononasiloxane. The base peak occurred at Si₂O (m/z 73). It constitutes 1.23% of the extract.

The chromatogram of the methanolic extract of the seeds of Moringa oleifera showed five peaks (Figure 4). These peaks (1-5) indicate the presence of five compounds (1-5) in the extract (Figure 2). The molecular formula, percentage composition and molecular mass of the compounds are shown in Table 2. These compounds comprise

mainly hydrocarbons, fatty acids, alcohols and esters. The constituent of the extract comprises; oleic acid (84%), L-(+) -ascorbic acid- 2, 6-dihexadecanoate (9.80%) 9-octadecenoic acid (1.88%), methyl esterhexadecanoic acid (1.31%) and 9-octadecenamide (0.78) (Table 2).

Compound 1 was identified as methyl ester-hexadecanoic acid with molecular formula of C17H34O2 (m/z 270) and base peak at m/z 74 which was due to loss of benzyl group ((C6H13) from the parent molecule. It constitutes 1.31% of the extract. Compound has molecular formula C38H68O8 (m/z 652) and constitutes 9.80% of the extract. The base peak occurred at C3H5O2 (m/z 73). This peak occurred due to McLafferty re-arrangement. Other prominent peaks observed on the compound occurred at m/z 43 (C3H7 +) and m/z 41 (C3H5). These peaks occurred due to proton migration and rearrangement. Compound 2 was identified as L-(+) - Ascorbic acid- 2, 6-dihexadecanoate. 3 has molecular Compound C19H36O2 (m/z 296) and base peak at m/z 55. The compound was identified as methyl ester 9-octadecenoic acid constituting 1.88% of the extract. Compound 4 is oleic acid with molecular formula C18H34O2 (m/z 282) and base peak at m/z 55. This constituent made up of 84% of the extract. Compound 5 was identified 9-octadecenamide with molecular formula C18H35NO (m/z 281) and base peak at m/z 59. The base peak occurred as a result of the detachment of C3H9 (m/z 57) and hydrogen molecule fragment from the compound. It constitutes 0.78% of the extract.

DISCUSSION

The methanolic extract of the leaves of *Moringa oleifera* showed sixteen peaks from the Gc-ms chromatogram. These peaks indicate the presence of sixteen compounds (1-16) in the extract (Figure 3). The composition of the extract comprises; 9-

Octadecenoic acid (20.89%), L-(+) -Ascorbic acid- 2, 6-dihexadecanoate (19.66%), 14 methyl -8-Hexadecenal (8.11%). hydroxyl-4-methyl-2-pentanone (7.01%), 3ethyl-2, 4-dimethyl-pentane (6.14%) and phytol (4.25%) as the major chemical constituents (Table 1). The flavouring phytochemical 2-pentanone reduces prostaglandin production COX-2 and expression in colon cancer cells. Inflammation and subsequent elevation of the enzyme cyclooxygenase-2 (COX-2) are two such factors involved in the development of colon cancer, and inhibition of these processes could be important targets for chemoprevention¹³. Organosulphur compounds (OSCs) prevent or slow down the carcinogenic process induced by a variety of carcinogens¹⁶. chemical OSCs protection against cancer. These include inhibition of the carcinogens, dermatitis and other minor wounds³. The occurrence of thiobenzoic acid and L-(+) -ascorbic acid 2, 6-dihexadecanoate in the leaves of M. oleifera may be the reason behind the use of the extracts in the treatment of wounds in herbal medicine in Nigeria². Ascorbic acid in the body helps in absorption from the intestine². It is required for connective metabolism especially the tissues, bones and teeth². It is necessary as anti-stress and protects against colds, chills and dumps. It prevents muscle fatigue and scurvy which is characterized by hemorrhages, bleeding gums, fragile bones, anemia and pains in the joints and defects in skeletal calcification 10. This function of ascorbic acid also accounts for its requirement for normal wound healing¹⁰. The hypothesis also supports the use of M. oleifera in treating wounds by the native communities in Nigeria. Ascorbic acid and OSCs act as antioxidants in the skin by scavenging and quenching free radicals generated by ultraviolet (UV) radiation from stabilization. Ascorbic acid and other phenolic compounds identified are important antioxidants. They act as electron

donors for eight important enzymes in humans². Ascorbic acid may protect against the oxidative damage of light in the eye and may also play an important role in sperm maturation¹¹. It helps in stabilizing plasma components and has been shown to be an effective scavenger of superoxide radical anion (H₂O₂), the hydroxyl radical (OH₂), singlet oxygen (O') and reactive nitrogen oxide (NO)¹¹.

The methanolic extract of the seeds of Moringa oleifera showed five peaks from the chromatogram (Figure 4). These peaks indicate the presence of five compounds (1-5). These compounds comprise mainly hydrocarbons, fatty acids, alcohols and esters. The composition of the extract comprises; oleic acid (84%) L-(+) - Ascorbic acid- 2, 6dihexadecanoate (9.80%) 9-octadecenoic acid (1.88%), methyl ester-hexadecanoic acid (1.31%) as the major constituents. The presence of fatty acids and their derivatives in oleifera seed extract dictates the pharmacological properties of the plant. Fatty acids and alcohols in the plant undergo esterification reaction to form esters². One or both of the oxygen atoms of carboxylic acid can be replaced by sulphur giving a thio acid or dithio acid respectively. Thio acids react readily with alcohols to form thio-esters. Thio-esters play an important part in the break down and synthesis of lipids and steroids in living tissues. Carboxylic acids are transferred from one enzyme reaction to another as thioesters of the complex thiol, Co enzyme A (CoA-SH). The thio-ester of benzoic acid with Co-enzyme A is the form in which acetate enters the sequence of enzyme catalyzed reactions which results in the synthesis of fatty acids and glycerides¹¹.

The constituent compounds in methanolic extract are long chain aliphatic carboxylic acids, (saturated and unsaturated) and their derivatives including alcohols, aldehydes as well as benzene carboxylic acid esters and a steroidal compounds. It is

pertinent to identify the possible roles of these constituent compounds in the curative properties attributed to the plant by herbal medical practitioners. Oleic acid is an unsaturated fatty acid present in several plants and being unsaturated is considered as a healthy source of fat in the diet. Many uncommon (secondary metabolite) fatty acids are known to have antibacterial and antifungal properties²¹. Dodecanoic. tetradecanoic. hexadecanoic octadecanoic and oleic acids are among the fatty acids known to have potential antibacterial and antifungal properties²¹. Oleic acid has been found to be fungistatic against a wide spectrum of moulds and yeasts. For example, it was observed to cause a delay of 6-8 hours in the germination of fungal spores, and was also found to be effective at low concentrations²¹. It has also been proposed that these fatty acids have antibacterial antifungal potential and principles application¹¹. for clinical Triterpene-fatty acid esters and free fatty including long chain C16-C20 unsaturated fatty acids were suggested to be responsible for the anti-inflammatory activity in the extract from M. oleifera seed²¹.

CONCLUSION

GC-MS analysis showed that there are 9-octadecenoic acid (20.89%), L-(+)- ascorbic acid- 2,6-dihexadecanoate(19.66%), 14 methyl -8-Hexadecenal (8.11%) , 4hydroxyl-4-methyl-2-pentanone (7.01%), 3ethyl-2, 4-dimethyl-pentane (6.14%), phytol (4.25%), octadeamethyl-cyclononasiloxane (1.23%), 1, 2-benzenedicarboxylic acid (2.46%), 3, 4-epoxy- ethanone comprising N-(-1-methylethyllidene)-benzene (1.78%). ethanamine (1.54%), 4. 8. tetramethylheptadecan-4-olide (2.77%), 3-5bis (1, 1-dimethylethyl)-phenol (2.55%), 1hexadecanol (1.23%), 3, 7, 11, 15hexadecene-1ol tetramethyl-2 (1.17%),hexadecanoic acid (2.03%) and 1, 2, 3propanetriyl ester-9 octadecenoic (1.23%) as the chemical composition in the leaf extract and oleic acid (84%) L-(+)ascorbic acid- 2, 6-dihexadecanoate (9.80%), 9-octadecenoic acid (1.88%), methyl esterhexadecanoic acid (1.31%)octadecenamide (0.78) are the chemical components in the seed extract. The study also revealed that 9-octadecenoic acid (20.89%) constitutes the major of the leaf extract while oleic acid (84%) is the major component of the seed extract. GC-MS analysis showed the presence of many chemical constituents of Moringa oleifera leaves and seeds locally grown in Abakaliki, Nigeria. The presence of various bioactive compounds confirms the application of Moringa oleifera for various ailments by traditional practitioners.

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REFERENCES

- 1. P.M. Aja, N. Nwachukwu, A. U. Ibiam, I. O. Igwenyi and P.N. Onu (2014). Comparative Evaluation of Transaminases and Alkaline Phosphatase Activities in Albino Rats Administered Aqueous, Ethanolic and Methanolic Extracts of Moringa oleifera Seeds Locally Grown in Abakaliki, Nigeria, *Journal of Biological and Chemical Research*, Volume 31 (1):164-181.
- 2. Akinmoladum, A. C., Ibukun, E.O., Akinsinlola, B. L., Onibon, T. R., Rinboboye, A. O., Obuofor, E. M. and Farombi, E. O. (2007). Chemical Constituents and Antioxidant Activity of *Alstonia boonei, African Journal of Biotechnology*, 6 (10): 1197-1201.
- 3. Brillo, A. J. A and Selvakymari, P. A. S., (2006). GC-MS Evaluation of Bioactive Compounds and Antibacterial Activity,

- Journal of Medicinal and Aromatic Plant Sciences, 26:578-579.
- 4. Dangi, S.Y, Jolly, C.I and Narayanan, S. (2002) Anti-hypertensive activity of the total alkaloids from the leaves of *Moringa oleifera*. *Journal of Pharmaceutient Biology*, (2): 144-148.
- 5. Ghosi, S., Nwobodo, E and Ofili, J. O., (2000). Hypocholesterolemic effect of crude extract of leaf of *Moringa oleifera* in high fat diet fed wistar rats. *Journal of Ethno pharmacology* 69 (1): 21-25.
- 6. Naik, G. H., Priyadarsini, K.I., Satav, J.G., Banavalikar, M. M., Sohani, D. P., Biyani, M. K. and Mohan, H. (2003). Comparative antioxidant activity of individual herbal components used in ayurvedic medicine. *Phytochemistry*, 63: 97-104.
- 7. Naznin, A, Mamunur, R and Amran M. S, (2008). Comparison of leaves extract with atenolol on serum triglyceride, serum cholesterol, blood glucose, heart weight, body weight in adrenaline induced rats. *Saudi Journal of Biological Sciences*, 15(2): 253-258.
- 8. Nikkon, F., Haque, M. E., Aragianis, K and Mosaddik, M. A. (2003). Isolation of aglycone of deoxy-nia-zimicin from *Moringa oleifera* and its cytotoxicity. *Rev. Latinoamur aunim* 31(1):5-9.
- Keay, R. J (1989). Medicinal Trees of Nigeria, 2nd edition. Oxford University Press, New York. 476-479.
- Okwu, D. E. and Emenike, I. N. (2006). *Moringa oleifera, International Journal of Molecular Medicine* Advance Science, 2 (1): 1-6.
- 11. Okwu, E. D. and Ighodaro, U. B., (2010). GC-MS evaluation of bioactive compounds and antibacterial activity of the oil fraction from the leaves of *Alstonia boonei* De Wild. *Der Pharma Chemica*, 2(1): 261-2.
- 12. Oyenuga, V.A. and Fetuga, B. L., (2003). Medical nutrient. Dietary important vegetables. 5th edition, Ibadan press. Nigeria, 122-129.
- 13. Petterson, J., Karlsson, P.G., Rafter, J. J., and Bohlin, L., (2008). The flavouring phytochemical 2-pentanone reduces prostaglandin production and COX-2

- expression in colon cancer cells, *Biological* and *Pharmaceutical Bulletin* 31:28-34.
- 14. Ranjan, R., Swamp, D., Patra, C. R., and Vikas, C. (2009). *Tamarindus indica* L. and *Moringa oleifera* extracts administration ameliorates fluoride toxicity in rabbits, *Indian Journal of Experimental Biology*, 49:900-905.
- 15. Rao, K. S and Mishra, S. H. (1998). Anti inflammatory and anti-hepatotoxic activities of fruits and the roots of *Moringa pterygosperma*, *Indian Journal of Pharmaceutical Science* 60:12-21.
- 16. Siess, M. H., Lebon, A. M., Teyssier, C., Belloir, C., Singh, V. and Berges, R. (2007). Medicinal and aromatic plant, *Science and Biotechnology*, 1(1): 25-30.
- 17. Valiathan, M. S. (1998) Healing plants. *Current Science*, 75 (11): 1122-1127.
- 18. Ogunlesi, M., Okiei, W and Osibote, A. E., (2010) Analysis of the essential oil from the

- leaves of *Sesamum radiatum*, a potential medication for male infertility factor, by gas chromatography-mass spectrometry, *African Journal of Biotechnology* 9(7), 1060-1067.
- 19. Sharma, V., Sharma, A and Kansal, L, (2010). The effect of oral administration of *Allium sativum* extracts on lead nitrate induced toxicity in male mice. *Food Chemistry and Toxicology*, 48: 928-936.
- World Health Organization (2003). Report WHO Health Benefit of Herbs. The Past, the present, the future. *Medicinal Journal of* USA, 182(4): 1-10.
- 21. Ogunlesi, M., Okiei, W and Osibote, A. E., (2010) Analysis of the essential oil from the leaves of *Sesamum radiatum*, a potential medication for male infertility factor, by gas chromatography-mass spectrometry, *African Journal of Biotechnology* Volume 9(7), 1060-1067.

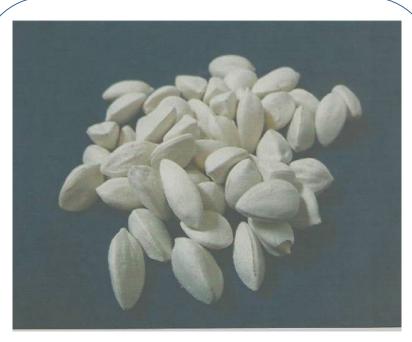


Figure 1. Dried seeds of Moringa oleifera

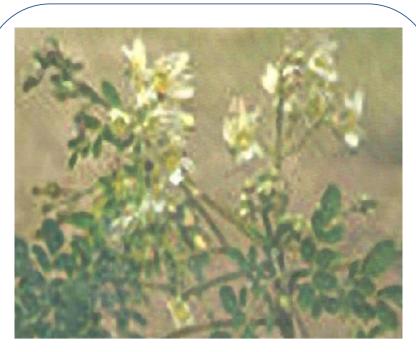
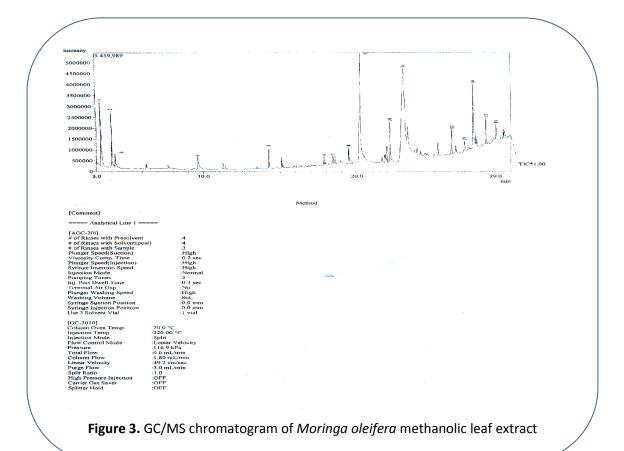


Figure 2. Moringa oleifera leaves



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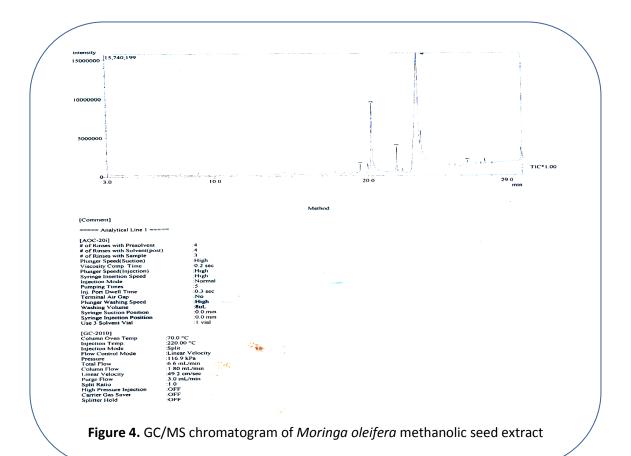


Table 1. GC-MS analysis and mass spectral data of methanolic fraction from the leaves of *Moringa oleifera* showing molecular formula, molecular weight, percentage content, retention time and base peak

Peak	Compound	Molecular formular	Molecular weight	Retention time	Percentage content	Mass peaks
1	4-hydroxy-4-methyl-2-pentane	C ₆ H ₁₂ O ₂	116	3.29	7.01%	42
2	3-ethyl-2,4-dimethyl-pentane	C ₆ H ₁₂ O	100	4.008	6.14%	49
3	3-4-epoxy-ethanone	C ₉ H ₂ 0	128	4.233	1.78%	35
4	N-(1-methylethyllidene)-benzene ethanamine	C ₁₁ H ₁₅ N	161	9.635	1.54%	50
5	3,5-bis(1,1-dimethylethyl)-phenol	C ₁₄ H ₂₂ O	206	14.250	2.55%	94
6	1-Hexadecanol	C ₁₆ H ₃₄ O	242	17.850	1.23%	64
7	3,7,11,15-Tetramethyl-2 hexadecene-1-ol	C ₁₆ H ₃₂ O	240	18.425	1.17%	67
8	Hexadecanoic acid	C ₁₇ H ₃₄ O ₂	270	19.458	2.03%	90
9	L-(+)-Ascorbic acid 2,6- dihexadecanoate	C ₃₈ H ₆₈ O ₈	652	20.183	19.66%	136
10	Phytol	C ₂₀ H ₄₀ O	296	22.142	4.24%	83
11	9-Otadecenoic acid	C ₁₈ H ₃₄ O ₂	282	23.000	20.89%	129
12	4,8,12,16-Tetramethyl heptadecan- 4-olide	C ₂₁ H ₄₀ O ₂	324	26.133	2.77%	172
13	9-Octadecenoic acid- 1,2,3-propanetrieyl ester	C ₅₇ H ₁₀₄ O ₆	884	26.983	1.23%	123
14	14-methyl-8-hexadecenal	C ₁₇ H ₃₂ O	252	27.533	8.11%	222
15	1,2-Benzene dicarb oxylic acid,	C ₂₄ H ₃₈ O ₄	390	28.358	2.46%	144
16	Octadecamethyl –cyclononasiloxane	C ₁₈ H ₅₄ O ₉ Si ₉	666	9.017	1.23%	199

Table 2. GC-MS analysis and mass spectral data of methanol fractions from the seeds of *Moringa oleifera* showing molecular formula, molecular weight, percentage content, retention time and base peak

Compound	Molecular formula	Molecular weight	Retention time	Percentage content	Base peak
Methyl ester- hexadecanoic acid	C ₁₇ H ₃₄ O ₂	270	19.458	1.31%	74
L-(+)-ascorbic acid 2, 6dihexa-decanoate	C ₃₈ H ₆₈ O	242	20.23	9.80%	73.05
Methyl ester-9- octadecenoic acid	C ₁₉ H ₃₄ O ₂	296	21.875	1.88%	55.05
Oleic acid	C ₁₅ H ₂₈ O ₂	240	23.233	84%	55.05
9-octadecenamide	C ₁₈ H ₃₅ NO	281	26.417	0.78%	59